

# Phosphate-Limited Medium with 100mM Ethanol as a Sole Source of Carbon and Energy

Nikolai Slavov

## 1 ORIGIN

This protocol describes the composition of the growth medium used by Slavov and Botstein (2011, 2010) for studying the growth rate response and by Slavov *et al* (2011) to discover metabolic cycling without cell division cycling. It is based on the composition of the medium described by Saldanha *et al* (2004) and later in the protocols of the Dunham and Botstein labs. It contains  $[KH_2PO_4] = 20mg/L$  as the only source of phosphorous and 100mM ethanol as the only source of carbon and energy. We have demonstrated that in this medium the growth of budding yeast is limited on phosphate and both the final biomass of batch cultures and the steady-state biomass density of continuous cultures depend linearly on the concentration of phosphate Slavov and Botstein (2011, 2010). The experiments used in optimizing the composition of this medium can be found at: [http://genomics-pubs.princeton.edu/grr/Experimental\\_Design.html](http://genomics-pubs.princeton.edu/grr/Experimental_Design.html)

## 2 COMPOSITION OF THE MEDIUM

Chemical	Amount
10× Salts, see Table 2	1.0 L
1000× metals, see Table 3	10.0 mL
1000× vitamins, see Table 4	10.0 mL
Ethanol, 95%	61.3 mL
10g/L $KH_2PO_4$ (monobasic)	20.0 mL
MilliQ Water	fill up to 10L

**Table 1.** Composition of the medium.

## 2.1 SALTS

Salts are made as a 10× stock that keeps at room temperature for at least a year, see Table 2. Make the salts in sterile glass with distilled water. Be vigilant about shaking before using.

Amount	Mineral Salt	Storage
5 g	$CaCl_2 \cdot 2H_2O$	RT shelf
5 g	$NaCl$	RT shelf
25 g	$MgSO_4 \cdot 7H_2O$	RT shelf
50 g	$KCl$	RT shelf
250 g	$(NH_4)_2SO_4$	RT shelf
fill up to 5L	MilliQ Water	Filtration System

**Table 2.** Mineral salts used for making 10× stock solution of the salts.

## 2.2 METALS

Metals are made as a 1000× stock that keeps at room temperature for at least a year, see Table 3. Keep the bottle well wrapped in foil since some of the metals are light sensitive. Make the metals in sterile glass with distilled water. Be vigilant about shaking before using since the metals will not totally dissolve. Dissolve the mineral salts in Table 3 in 900mL distilled water, in stirring glass.

Amount	Metal Salt	Storage
500 mg	boric acid	RT shelf
40 mg	copper sulfate.5H <sub>2</sub> O	RT shelf
100 mg	potassium iodide	RT, dark, dessicator
200 mg	ferric chloride.6H <sub>2</sub> O	RT shelf
400 mg	manganese sulfate.H <sub>2</sub> O	RT shelf
200 mg	sodium molybdate.2H <sub>2</sub> O	RT shelf
400 mg	zinc sulfate.7H <sub>2</sub> O	RT shelf

**Table 3.** Mineral salts used for making 1000× stock solution of the metals.

Bring total volume to 1L with MilliQ water, and pour into a bottle. Cover the bottle with foil, and store at room temperature.

## 2.3 VITAMINS

Vitamins are also made as a 1000× stock, see Table 4. The solution is aliquoted into 50ml Falcon tubes and stored at  $-20^{\circ}\text{C}$ . Don not fill the tubes to the top, or else the lid will split when frozen. The “working tube” can be stored at  $4^{\circ}\text{C}$ . The vitamins will not dissolve completely, so shake before use. Care should be taken to keep the solution well mixed while aliquoting. Weight all chemicals and add to a beaker of stirring glass distilled water to dissolve as much as possible. Top off to 1L, then aliquot about 40mL per 50mL tube, and freeze.

Amount	Vitamin	Storage
2 mg	biotin	$4^{\circ}\text{C}$
400 mg	calcium pantothenate	$4^{\circ}\text{C}$
2 mg	folic acid	RT, dark, dessicator
2000 mg	inositol (aka myo-inositol)	RT shelf
400 mg	niacin (aka nicotinic acid)	RT shelf
200 mg	p-aminobenzoic acid	$4^{\circ}\text{C}$
400 mg	pyridoxine HCl	RT, dark, dessicator
200 mg	riboflavin	RT shelf
400 mg	thiamine HCl	RT, dark, dessicator

**Table 4.** Vitamins used for making 1000× stock solution of the vitamins

## References

- Saldanha AJ, Brauer MJ, Botstein D (2004) Nutritional Homeostasis in Batch and Steady-State Culture of Yeast. *Mol Biol Cell* **15**: 4089–4104
- Slavov N, Botstein D (2010) *Universality, specificity and regulation of S. cerevisiae growth rate response in different carbon sources and nutrient limitations*. Ph.D. thesis, Princeton University
- Slavov N, Botstein D (2011) Coupling among growth rate response, metabolic cycle, and cell division cycle in yeast. *Mol Biol Cell* **22**: 1997–2009

Slavov N, Macinskas J, Caudy A, Botstein D (2011) Metabolic cycling without cell division cycling in respiring yeast. *Proceedings of the National Academy of Sciences of the United States of America* **108**: 19090–19095, PMID: 22065748